

## TOTAL SYNTHESSES OF ( $\pm$ )-TELEOCIDIN B-3 AND B-4<sup>1)</sup>

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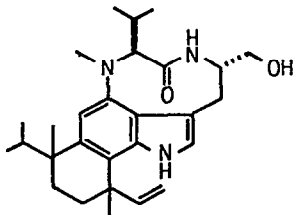
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Abstract; First total syntheses of ( $\pm$ )-Teleocidin B-3(1c) and B-4(1d) were achieved by several functionalizing reactions starting from indole.

Teleocidin B was first isolated as a strong skin-irritant compound from the mycelium of *Streptomyces mecliocidicus* by Takashima and Sakai.<sup>2)</sup> Recently, Fujiki *et al.*<sup>4)</sup> reported that teleocidin B is one of the most potent tumor promoters known. The chemical structure was determined by Hirata's group<sup>3)</sup> by chemical methods and X-ray analysis of the monobromoacetate of its dihydro derivative in 1966. Fujiki and Sakai's group<sup>5)</sup> found that the compound was a mixture of four stereoisomers [B-1(1a), B-2(1b), B-3(1c), and B-4(1d)] and that Hirata's teleocidin B derivative used for X-ray analysis was derived from teleocidin B-4. Teleocidin B is a unique natural product having an amino group at the 4-position and a modified mono terpenyl moiety at the 6,7-positions of the indole nucleus.

We have developed new functionalizing methods<sup>6,7)</sup> for the benzene part of indole derivatives and applied them to the efficient syntheses of indolactam v<sup>8)</sup> (the minimum structure for biological activity of teleocidin) and an indole derivative<sup>9)</sup> containing the same substituents at the 6,7-positions of teleocidin B. Now we report the first total syntheses<sup>10)</sup> of teleocidin B-3(1c) and B-4(1d).

We prepared benzoate 3<sup>9)</sup> by radical oxidation of 2 with benzoyl peroxide in 30% yield. Selective nitration<sup>11)</sup> of 3 was achieved by treatment with AcONO<sub>2</sub> in acetic anhydride. Other nitration conditions (for instance, conc. HNO<sub>3</sub> in acetic acid) could not be used because of instability of the product in such



Teleocidin B (1)  
[ mixture of B-1(1a), B-2(1b), B-3(1c), and B-4(1d) ]

acidic conditions. The position of the nitro group was established by comparison of the chemical shift of the 5-proton with those of the other 4-nitro derivatives.<sup>11)</sup> The benzoyl group of the nitro compound was selectively hydrolyzed with 1 N NaOH to give 4<sup>12)</sup> in 60% overall yield from 3. Oxidation of 4 with MnO<sub>2</sub> in CH<sub>2</sub>Cl<sub>2</sub> at 25°C for 1.5 hr afforded a lactam 5<sup>12)</sup> in 74% yield. Although the lactam 5 was a mixture of trans- and cis-isomers (5a:5b = 5:1), the former (5a), mp 178.5-180 °C, was obtained in pure form by crystallization.

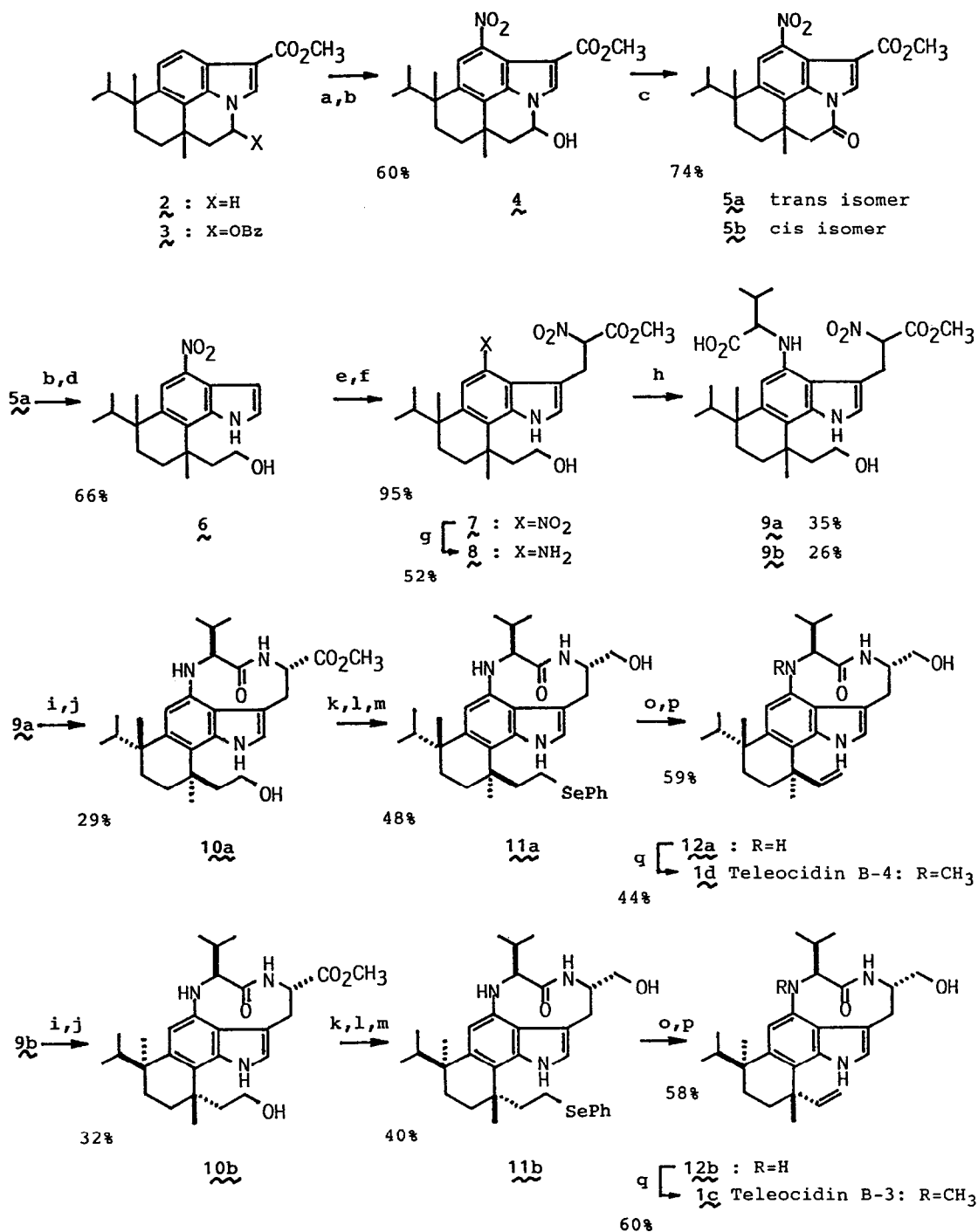
Interestingly, hydrolysis of 5a with 6 N NaOH at 60°C was accompanied by decarboxylation and subsequent reduction of the product with BH<sub>3</sub> in THF at 25 °C for 20 min afforded an alcohol 6<sup>12)</sup> in 66% yield. Compound 6 was treated with formalin and dimethylamine in acetic acid at 60°C for 15 min and followed by treatment with methyl nitroacetate in the presence of Et<sub>3</sub>N in chlorobenzene at 100°C for 10 min to give 7 (95%, 1:1 mixture of stereoisomers).

Selective reduction of the 4-nitro group of 7 by catalytic hydrogenation was unsuccessful, unlike our indolactam V synthesis,<sup>8)</sup> but reduction with sodium dithionite in the presence of Et<sub>3</sub>N in MeOH-H<sub>2</sub>O gave the corresponding 4-amino compound 8 in 52% yield. Introduction of a C<sub>5</sub> unit to 8 was achieved with 2-oxoisovaleric acid in DMF and subsequent reduction with NaBH<sub>3</sub>CN to give 9. Compound 9 was a mixture of four stereoisomers and could be easily separated on a silica gel TLC plate into two mixtures containing two stereoisomers, 9a<sup>12)</sup> (35%, 1:1 mixture) and 9b<sup>12)</sup> (26%, 1:1 mixture). Presumably these isomers are epimers related to the stereochemistry between the valinyl moiety and the terpenyl substituent at the 6,7-position because of the interaction of those substituents on the very flat tetrahydrobenzindole system. These stereochemistries were confirmed by the subsequent derivations to teleocidin B-3 and B-4.

Compound 9a gave a lactam 10a<sup>12)</sup> in 29% yield by the treatment with N-hydroxysuccinimide (HOSU) and dicyclohexylcarbodiimide (DCC) in CH<sub>3</sub>CN at 25°C for 30 min followed by H<sub>2</sub>/PtO<sub>2</sub> in methanol for 15 min. The epimer of 10a was not isolated.

Transformation of the hydroxyethyl group of 10a to the terminal olefin found in teleocidin B was a very difficult process because this alcohol cyclized<sup>13)</sup> easily at the 1-position of the indole nucleus. We chose the cis-elimination method of selenoxide derivative. Treatment of 10a with MsCl in pyridine followed by NaSePh in ethanol to give a selenide. Reduction of the ester group of the selenide with NaBH<sub>4</sub>/LiCl in THF-EtOH gave 11a which was isolated on silica gel TLC in 48% yield. Oxidation of the phenylselenenyl group of 11a with KIO<sub>4</sub> in THF-MeOH followed by heating at 60°C gave terminal olefin 12a<sup>12)</sup> in 59% yield.

Methylation<sup>3g)</sup> of 12a was achieved with CH<sub>3</sub>I/ NaHCO<sub>3</sub>-Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> in EtOH at 70 °C in a sealed tube for 4.5 hr to give teleocidin B-4 (1d) in 44% yield (54% of 12a was recovered). In the same manner, we also prepared teleocidin B-3



a) AcONO<sub>2</sub> b) NaOH c) MnO<sub>2</sub> d) BH<sub>3</sub>·THF e) CH<sub>2</sub>=N<sup>+</sup>(CH<sub>3</sub>)<sub>2</sub>(OAc)<sup>-</sup> f) O<sub>2</sub>NCH<sub>2</sub>CO<sub>2</sub>CH<sub>3</sub> / Et<sub>3</sub>N g) Na<sub>2</sub>S<sub>2</sub>O<sub>4</sub> h) (CH<sub>3</sub>)<sub>2</sub>CHCOCO<sub>2</sub>H/NaBH<sub>3</sub>CN i) HOSu / DCC j) H<sub>2</sub> / PtO<sub>2</sub> k) MsCl/Py l) NaSePh m) NaBH<sub>4</sub>-LiCl o) KIO<sub>4</sub> p) 60 °C q) CH<sub>3</sub>I

(1c) from 9b in 4.5% overall yield. <sup>1</sup>H-NMR, MS, and UV spectra and retention times on HPLC of the synthetic (±)-teleocidin B-3 (1c) and B-4 (1d) were completely identical with those of natural teleocidin B-3 and B-4, respectively.

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## REFERENCES AND FOOTNOTES

1. Synthetic studies on teleocidin V. <sup>8</sup>)
2. M. Takashima and H. Sakai, *Agric. Biol. Chem.*, **24**, 647(1960).
3. H. Nakata, H. Harada, and Y. Hirata, *Tetrahedron Lett.*, 2125(1966). H. Harada, N. Sakabe, Y. Hirata, Y. Tomiie, and I. Nitta, *Bull. Chem. Soc. Japan*, **39**, 1773(1966).  
Related natural products were reported recently. a) Teleocidin A. <sup>2</sup>)  
b) Lyngbyatoxin A: J. H. Cardellina II, F. -J. Marner, and R. E. Moore, *Science*, **204**, 193(1979). c) Indolactam V: K. Irie, M. Hirota, N. Hagiwara, K. Koshimizu, H. Hayashi, S. Murao, H. Tokuda, and Y. Ito, *Agric. Biol. Chem.*, **48**, 1269(1984). d) Olivoretin A and D: S. Sakai, N. Aimi, K. Yamaguchi, Y. Hitotsuyanagi, C. Watanabe, K. Yokose, Y. Koyama, K. Shudo, and A. Itai, *Chem. Pharm. Bull.*, **32**, 354(1984). e) Olivoretin B and C: Y. Hitotsuyanagi, K. Yamaguchi, K. Ogata, N. Aimi, S. Sakai, Y. Koyama, Y. Endo, K. Shudo, A. Itai, and Y. Iitaka, *Chem. Pharm. Bull.*, **32**, 3774(1984). f) Blastmycetin A-C: K. Irie, N. Hagiwara, T. Kurome, H. Hayashi, M. Arai, and K. Koshimizu, *Agric. Biol. Chem.*, **51**, 285(1987). g) Olivoretin E: S. Sakai, Y. Hitotsuyanagi, K. Yamaguchi, N. Aimi, K. Ogata, T. Kuramochi, H. Seki, R. Hata, H. Fujiki, M. Suganuma, T. Sugimura, Y. Endo, K. Shudo, and Y. Koyama, *Chem. Pharm. Bull.*, **34**, 4883(1986).
4. H. Fujiki, M. Mori, M. Nakayasu, M. Terada, and T. Sugimura, *Biochem. Biophys. Res. Commun.*, **90**, 976(1979).
5. Y. Hitotsuyanagi, H. Fujiki, M. Suganuma, N. Aimi, S. Sakai, Y. Endo, K. Shudo, and T. Sugimura, *Chem. Pharm. Bull.*, **32**, 4233(1984).
6. a) S. Nakatsuka, H. Miyazaki, and T. Goto, *Tetrahedron Lett.*, **21**, 2817(1980). b) S. Nakatsuka, H. Miyazaki, and T. Goto, *Chemistry Lett.*, 407(1981). c) S. Nakatsuka, K. Yamada, and T. Goto, *Tetrahedron Lett.*, **27**, 4757(1986). d) S. Nakatsuka, O. Asano, and T. Goto, *Heterocycles*, **24**, 2109(1986). e) S. Nakatsuka, K. Ueda, O. Asano, and T. Goto, *Heterocycles*, **26**, 65(1987). f) S. Nakatsuka, O. Asano, K. Ueda, and T. Goto, *Heterocycles*, **26**, in press (1987). g) O. Asano, S. Nakatsuka, and T. Goto, *Heterocycles*, **26**, in press (1987). h) S. Nakatsuka, K. Teranishi, S. Hayashi, and T. Goto, *Tetrahedron Lett.*, **28**, in press (1987).
7. S. Nakatsuka, T. Masuda, O. Asano, T. Teramae, and T. Goto, *Tetrahedron Lett.*, **27**, 4327(1986).
8. S. Nakatsuka, T. Masuda, K. Sakai, and T. Goto, *ibid.*, **27**, 5735(1986).
9. S. Nakatsuka, T. Masuda, and T. Goto, *ibid.*, **27**, 6245(1986).
10. Synthetic studies toward teleocidins by other groups were cited in reference 7.
11. Synthetic studies on teleocidin III, T. Masuda, K. Ueda, O. Asano, S. Nakatsuka, and T. Goto, *Heterocycles*, **26**, in press (1987).
12. Typical proton signals of <sup>1</sup>H-NMR spectra in CDCl<sub>3</sub>, δ(ppm). 4(major isomer): 5.92(1H,br.m),7.59(1H,s),8.05(1H,s). 5a:0.73(3H,d,J=8),1.08(3H,d,J=8),1.28(3H,s),1.32(3H,s),2.77(1H,d,J=16),2.90(1H,d,J=16). 5b:0.80(3H,d,J=8),0.98(3H,d,J=8),1.28(3H,s),1.52(3H,s),2.85(2H,s). 6:2.20(2H,t,J=8),3.36(1H,m),3.68(1H,m),7.22(1H,dd,J=3 and 2),7.38(1H,t,J=3),8.13(1H,s). 9a:3.79 and 3.83(3H,s),3.93 & 3.94(1H,d,J=7). 9b:3.80 and 3.82(3H,s),3.98 and 4.00(1H,d,J=6). 10a:3.73(3H,s),5.12(1H,br.d,J=10),5.40(1H,br.d,J=10). 12a:5.24(1H,d,J=10),5.38(1H,d,J=18),6.14(1H,dd,J=18 and 10).
13. For instance, treatment of 7 with formic acid at 25°C afforded a cyclized product similar to 2.

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